

## Consider organisms developing in their natural environment

- What perturbations will the system encounter?
- What aspects of mechanics matter?
- How does mechanics contribute to phenotypic variation?



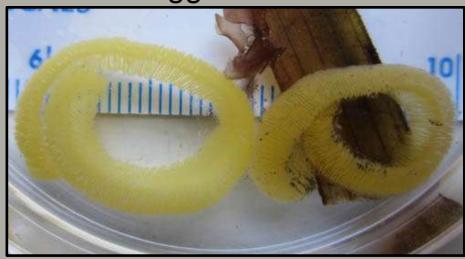
### Embryos in False Bay

#### Snail



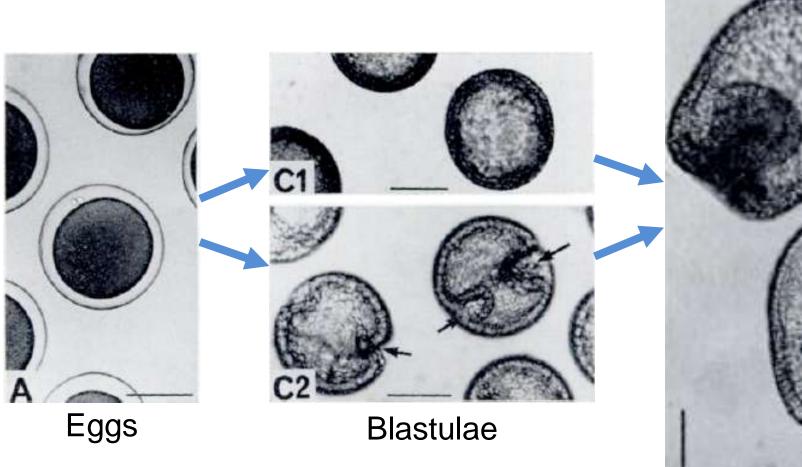


Egg masses



\*images from Yasmin von Dassow

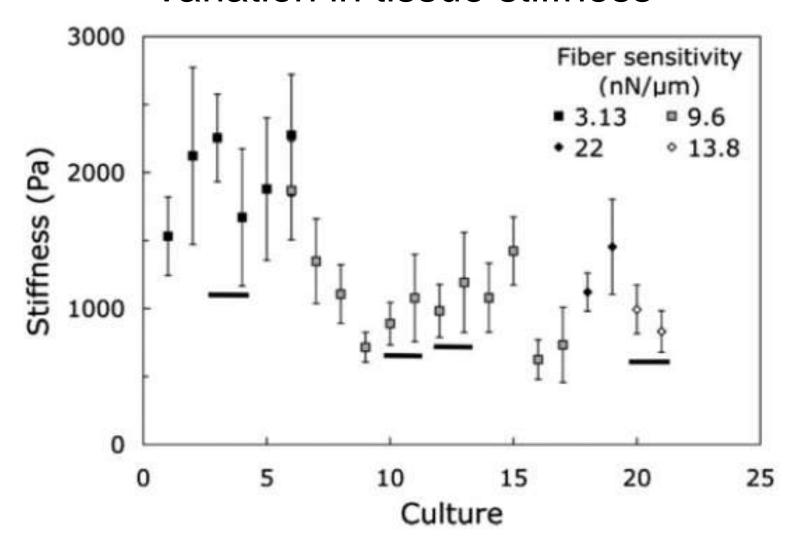
# Starfish embryos Salinity driven variation in embryo form



Kobayakawa, Y., Satoh, N., 1978. Induction of the wrinkled blastula formation in the starfish, *Asterina pectinifera*, by modified developmental conditions. Biological Bulletin 155, 150-160. Fig. 1

Gastrulae

## Sea urchin embryos Variation in tissue stiffness



von Dassow, M., Davidson, L.A., 2007. Birth Defects Res C Embryo Today 81, 253-269 (Based on data from Davidson, L.A., Oster, G.F., Keller, R.E., Koehl, M.A., 1999. Dev Biol 209, 221-238.)

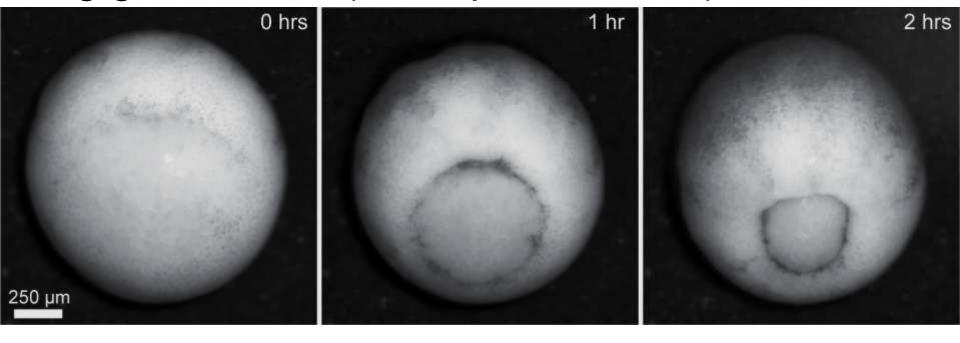
Is morphogenesis sensitive to tissue mechanics?

Natural variability sets lower bound on sensitivity

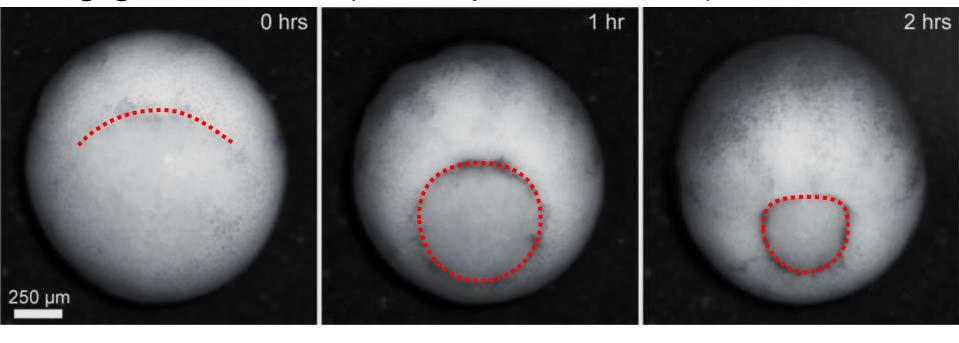




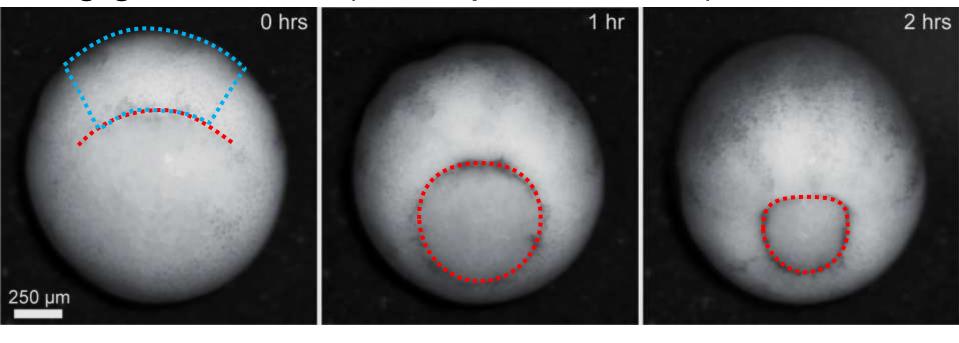
### Frog gastrulation (blastopore closure)



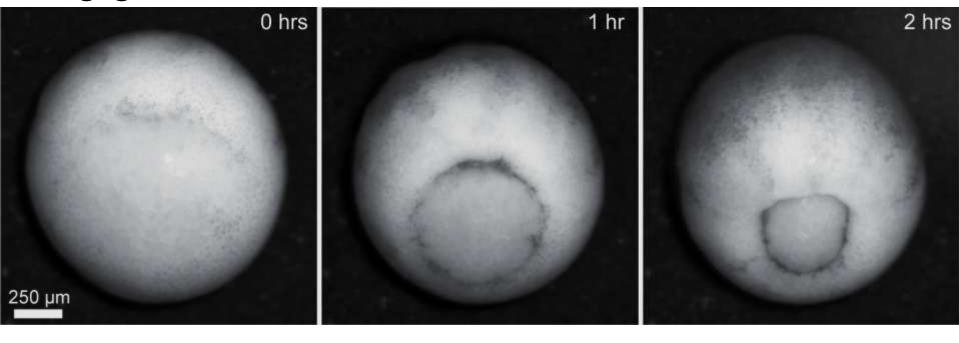
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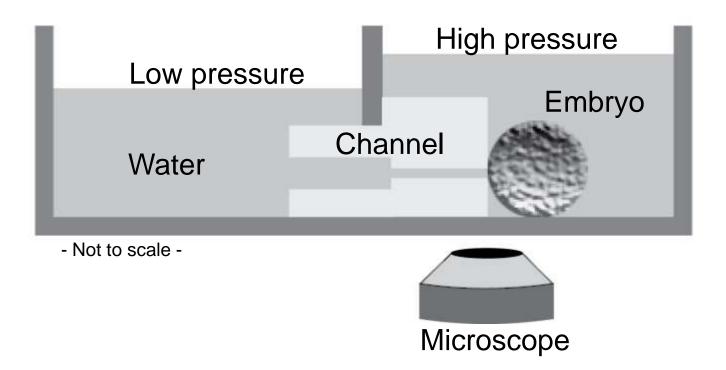
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### Frog gastrulation

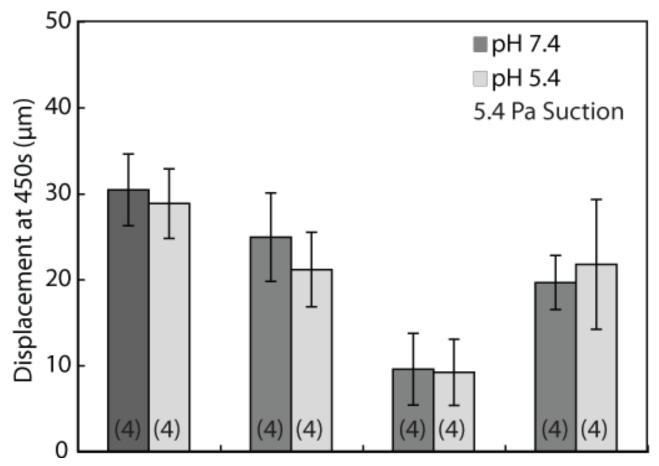


## Micro-aspiration: Measure mechanics with intact embryos



Displacement is inversely proportional to tissue stiffness

## How much do environmental factors contribute to mechanical variation?



- Neither pH nor salt concentration affected stiffness
- 0.3 µM Latrunculin B reduced stiffness\*
- High clutch to clutch variation\*

\**P* ≤ 0.05, 2-way ANOVA

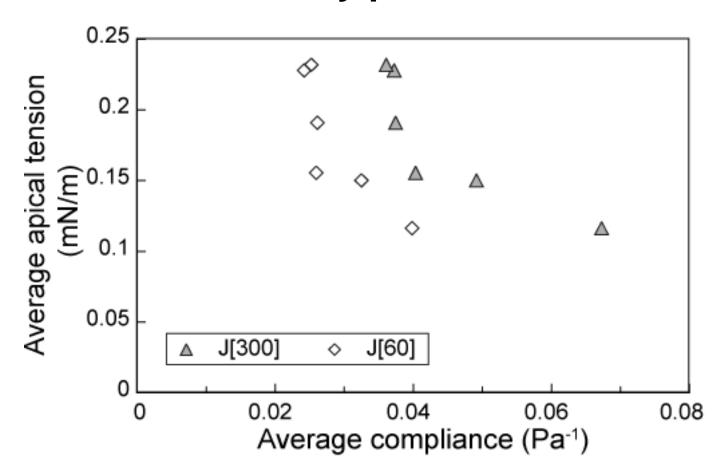
#### Conclusions (part 1):

- Natural variation in mechanics is substantial.
- Morphogenesis is surprisingly insensitive to mechanical variation.

What properties allow morphogenesis to be so robust?

Coupling between stiffness and force production?

#### Softer clutches may produce lower forces



J(60): P = 0.04; J(300):  $P \le 0.01$ ; Kendall's Tau

# Do changes in morphogenetic rates require fine control of tissue mechanics?

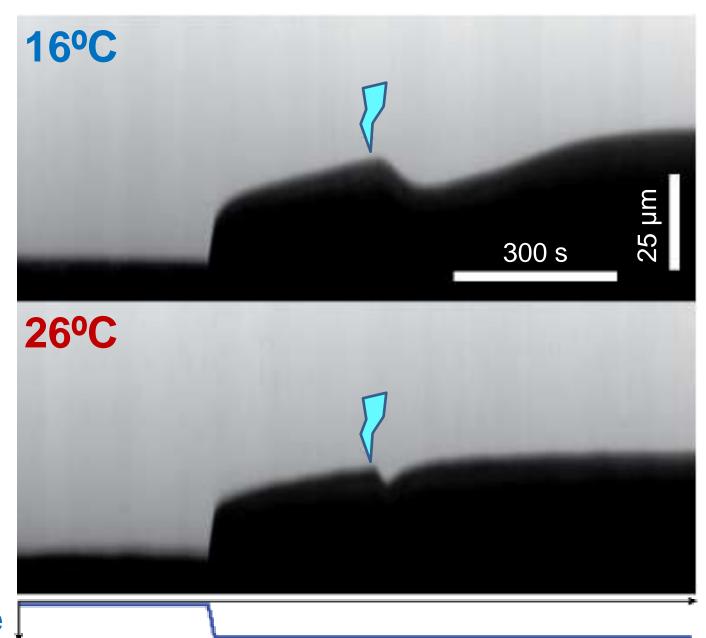
Morphogenesis is faster at high temperatures.

Therefore, we expect warm embryos to:

- Be softer and/or
- Exert more force

#### We expect warm embryos to:

- Be softer and/or
- Exert more force
- Compared embryos at 16° to 26°C
- Used micro-aspiration to determine viscoelastic properties
- Used induced contractions as proxy for force generation



Flectrical stimulation

Pressure

We expected warm embryos to:

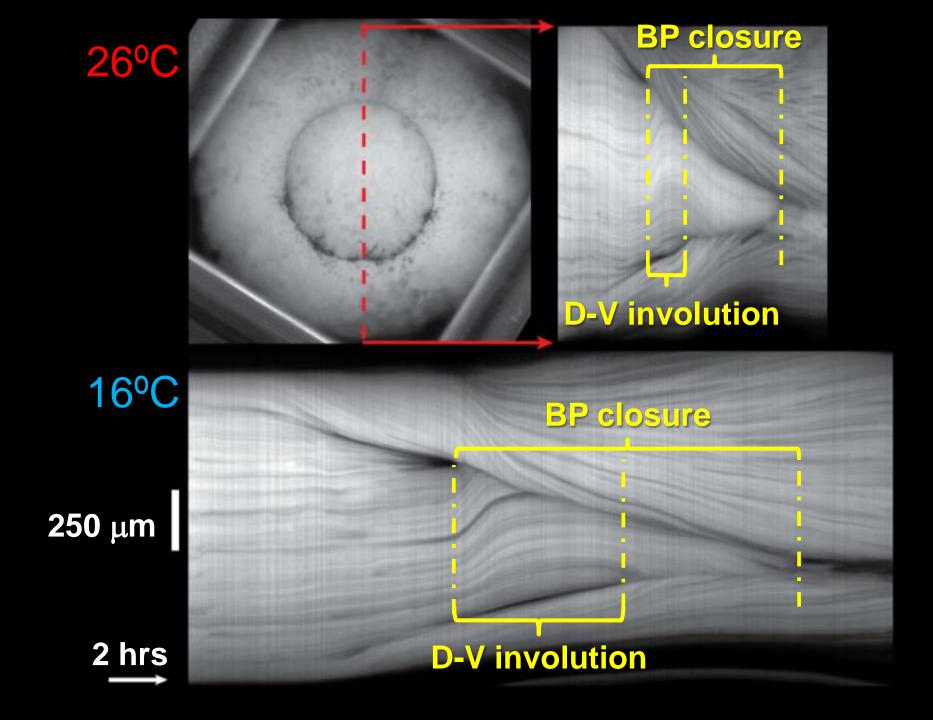
Be softer

and/or

Exert more force

Does morphogenesis change?

- Compared embryos at 16° to 26°C
- Measured relative timing of two processes



We expected warm embryos to:

Be softer

and/or

Exert more force

Does morphogenesis change?

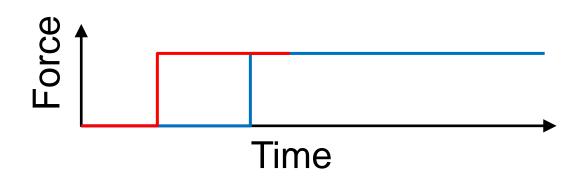
Yes

Could mechanics drive the temperaturedependent differences in morphogenesis?

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#### **Assumptions**

Forces:

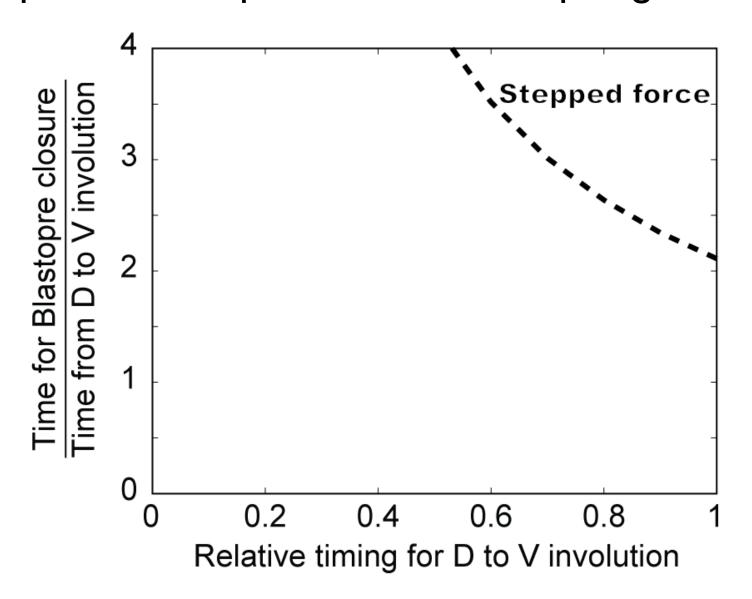


Viscoelasticity:

$$J(t) = J(1)^*t^{\beta} = J(1)^*t^{\beta}$$

Geometry:

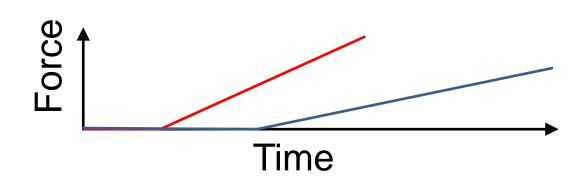
1D, small deformation



### Could mechanics drive the temperaturedependent differences in morphogenesis?

#### **Assumptions**

Forces:

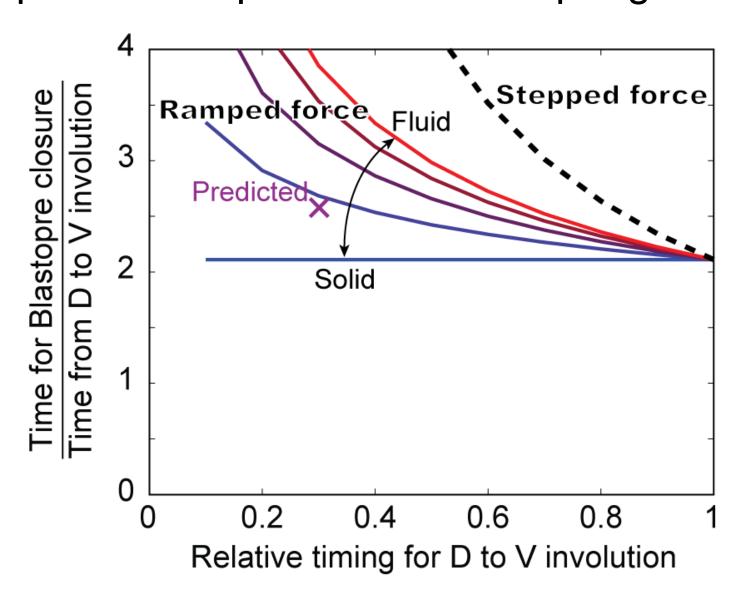


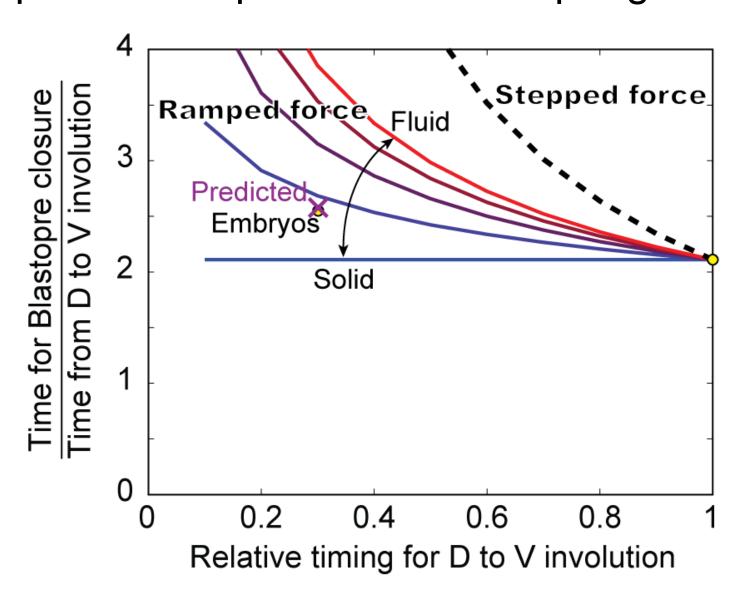
Viscoelasticity:

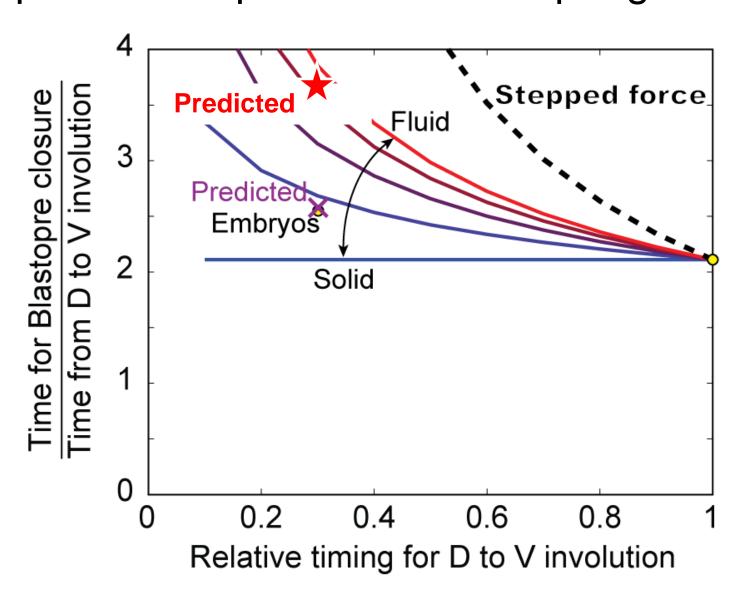
$$J(t) = J(1)^*t^{\beta} = J(1)^*t^{\beta}$$

Geometry:

1D, small deformation







### Conclusions (part 2)

- Organism-environment interactions suggest new ways to think about mechanics.
- Xenopus tolerates 2x range in tissue stiffness
- Xenopus tolerates >3x range in developmental rate due to:
  - Tolerance of morphogenetic variation
  - Not modifying tissue mechanics
- Mechanics affects variation in morphogenesis

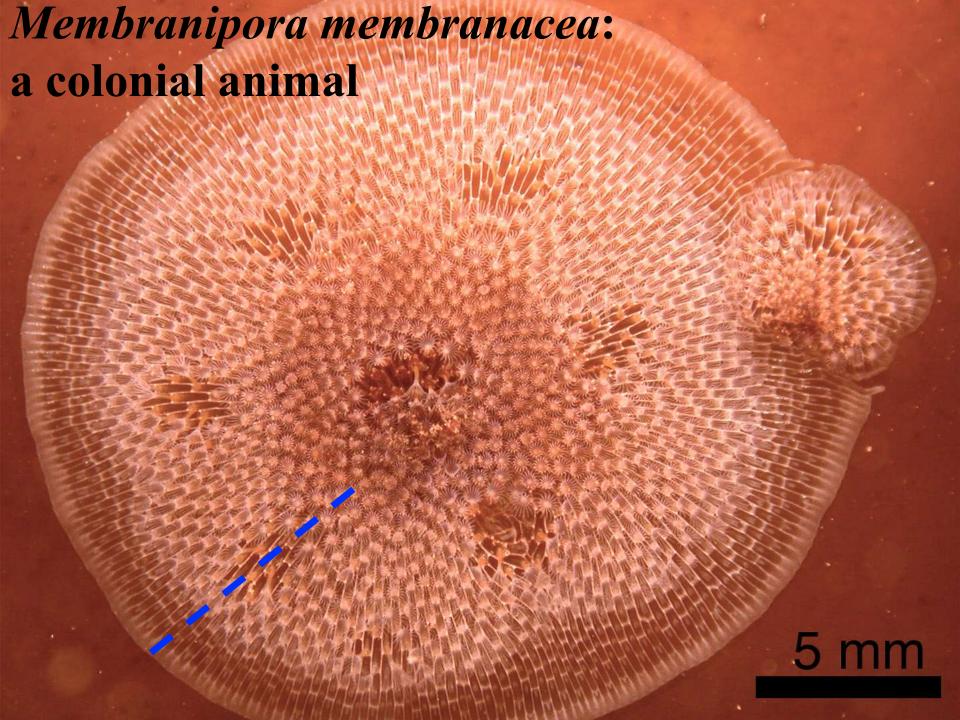


#### Thanks to:

#### Mimi Koehl

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Friday Harbor Labs
HHMI Predoctoral Fellowship
UC Berkeley Dept. of Integrative Biology



### A colony with a chimney, side view

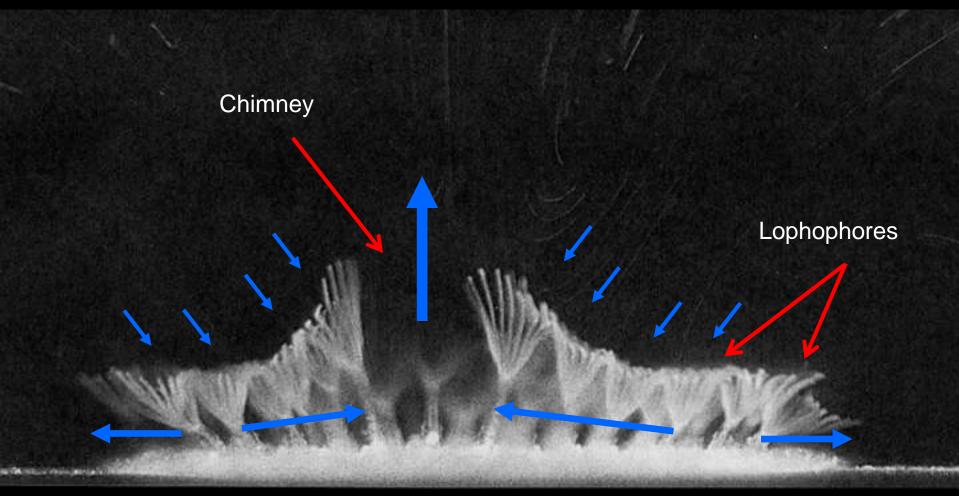
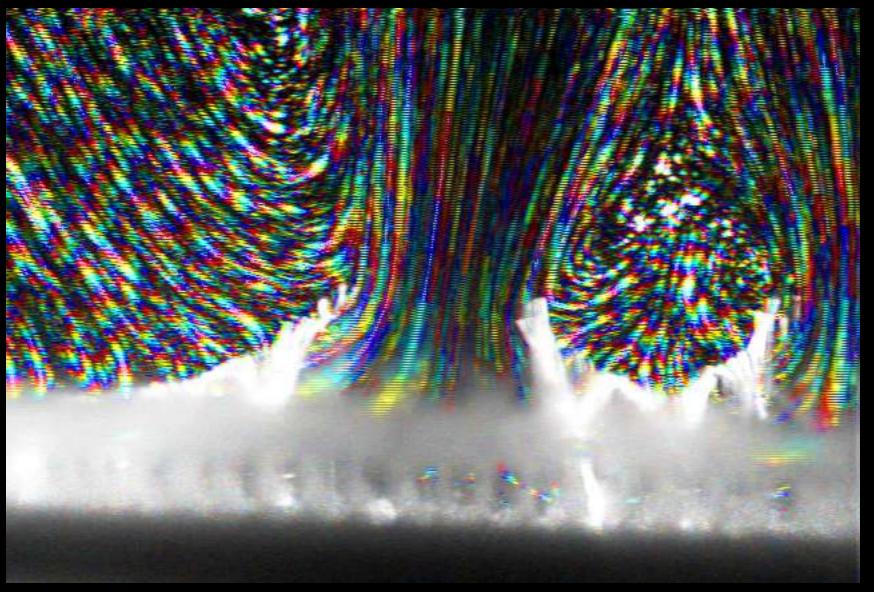
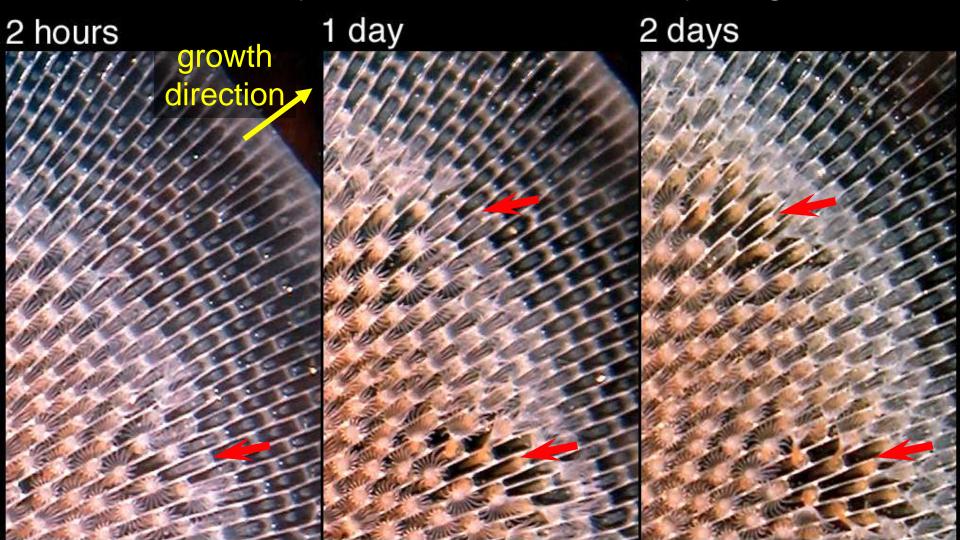


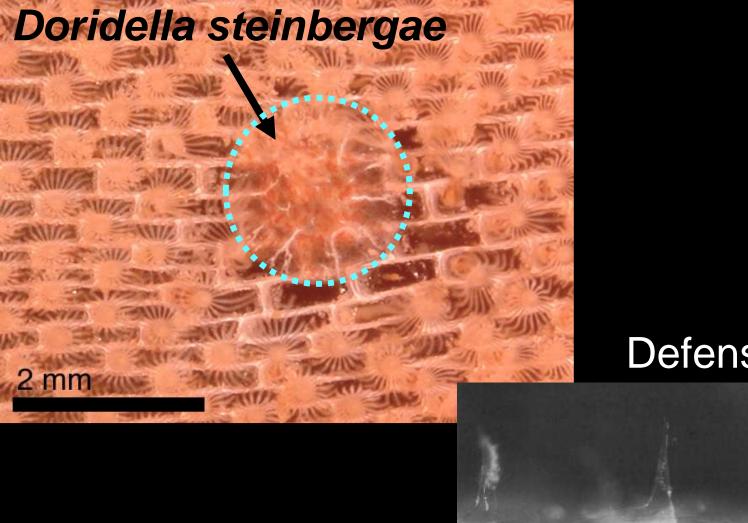
Photo from Grunbaum, D., 1997. Limnology and Oceanography 42, 741-752.

### Flow generated by colony



### Chimneys form at the canopy edge





Defensive spines

Photo from Grunbaum, D., 1997. Limnology and Oceanography 42, 741-752.

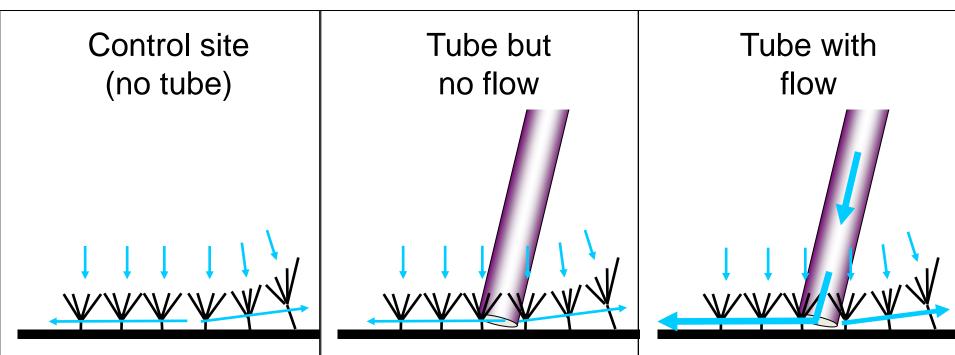
- Phenotypic plasticity
  - → Flow-regulated patterning?
- Predatory nudibranchs
  - → Defensive spines form
    - → Raise resistance to flow
      - → Reduce chimney spacing

• Chimney formation & zooid size injuries substrate shape

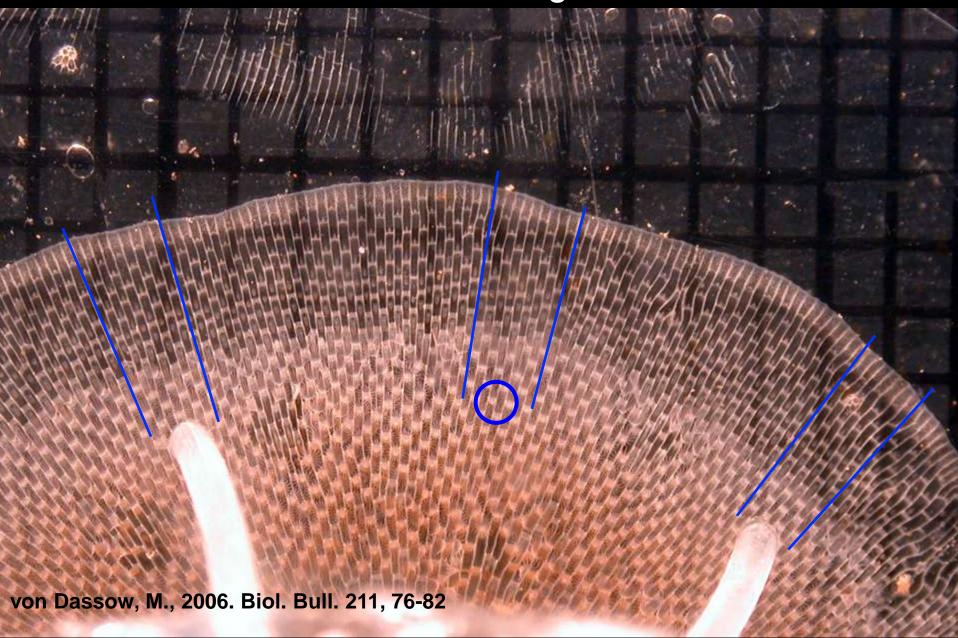
Does flow control where chimneys form?

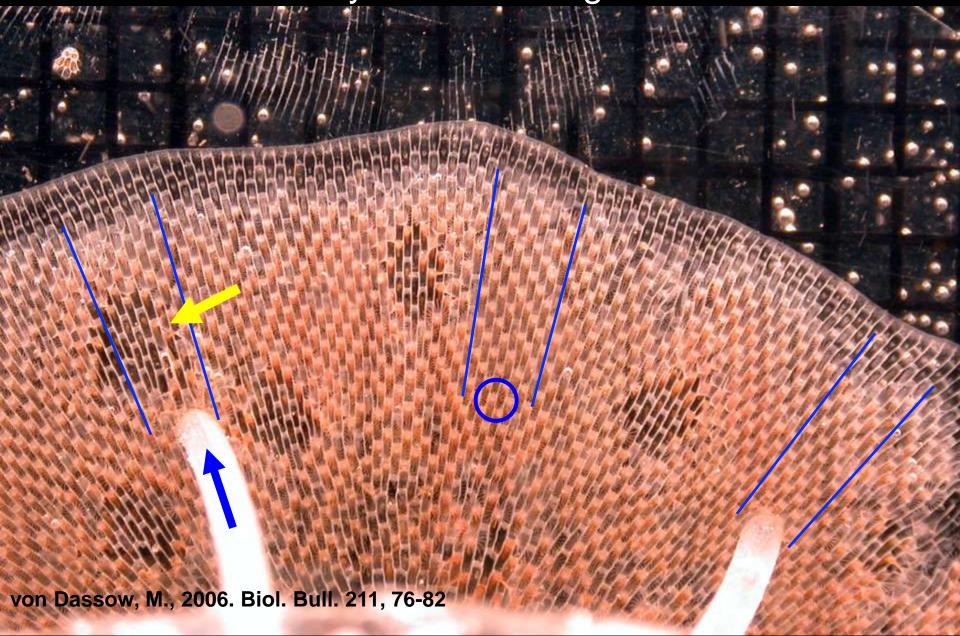
# Does high excurrent flow speed induce chimney formation?

Injected seawater under the canopy to increase the flow out the canopy edge



### Prior to starting flow





## Lophophores surrounding the opening are taller than neighbors n = 6 (out of 6)

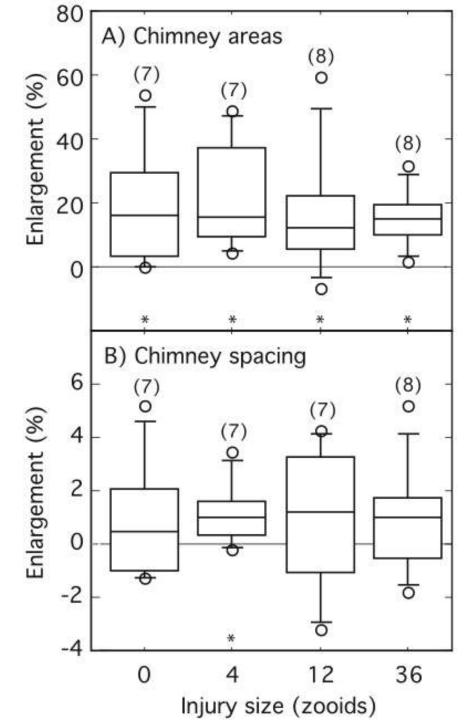


- High flow → Chimney formation
- Flow affects conduit formation in both internal and external fluid transport systems

How might this affect responses to the environment?

Injuries did not affect nearby chimneys

Perturbations do not spread to existing chimneys



von Dassow, M., 2005. Biol. Bull. 208, 47-59

#### Conclusions (part 3)

- Flow affects patterning in an external fluid transport system
  - → Drives responses to environment?
- But, changes in flow may not affect existing chimneys
  - → Limits responses to environment?

### Consider organisms in their environment

 What perturbations does the system normally face?

- What aspects of mechanics matter?
- How does mechanics contribute to phenotypic variation?

